## A New Diterpenoid Dimer from Annona glabra

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**Abstract**: A new diterpenoid dimer annonebinide A has been isolated from the stems of *Annona glabra*. Its structure was determined to be *ent*-16 $\alpha$ -hydroxykauran-17-yl *ent*-16 $\beta$ -kauran-17-oate on the basis of spectroscopic and chemical evidence.

Keywords: Annona glabra, diterpenoid dimer, ent-16α-hydroxykauran-17-yl ent-16β-kauran-17-oate.

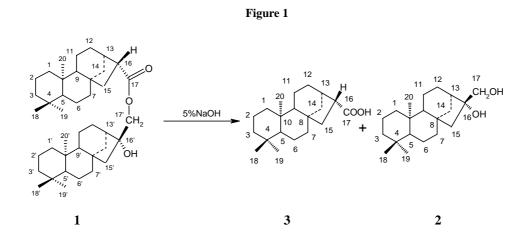
Annona glabra L. (Annonaceae), commonly known as "pond apple", is a tropical tree distributed mainly in the American and in southeast Asia. It has been used as an insecticide and parasiticide<sup>1-2</sup>. The genus Annona comprises about 120 species, many of them grow in tropical area in America and a few grow in tropical area of Africa. Five species were planted in China. Many plant of the genus Annona are excellent fruits in tropical area, but they are used as medicine in antivirus and antitumor in folk medicine<sup>3</sup>. Therefore it is necessary to study further the chemistry and pharmacology of Annona plants in order to clarify the relationship between folk medicines and their ingredients. In our studies, a series of kaurane diterpenes and other compounds were isolated from the bark of A. glabra. Among them, a new diterpenoid dimer was first found in the family of Annonaceae, named annonebinide A (1). In this communication we report its structure.

The air-dried stems (3.5 kg) of *A. glabra* were successively extracted with hot petroleum ether, EtOAc and EtOH. The EtOAc extract was fractionated by repeated silica gel column chromatography to yield annonebinide A.

Compound **1** was obtained as colorless needles (EtOAc), mp 275-277°C,  $[\alpha]_{D}^{25}$ -50 (c 0.25, CHCl<sub>3</sub>), HRFAB-MS showed a molecular formula of C<sub>40</sub>H<sub>64</sub>O<sub>3</sub>, *m/z* [M<sup>+</sup>] 592.4768 (calcd. 592.4858). Its IR spectrum showed the presence of a hydroxyl group (3413cm<sup>-1</sup>) and an ester carbonyl group (1703 cm<sup>-1</sup>). The <sup>1</sup>H-NMR spectrum showed signals due to six tertiary methyl groups at  $\delta$  0.80 (s, 6H), 0.84 (s, 6H), 0.99 (s, 3H) and 1.01 (s, 3H), one oxymethylene group at  $\delta$  4.02 (d, 1H, J=11.3Hz) and 3.86 (d, 1H, J=11.3Hz) (**Table 1**). The signals at chemical shifts of  $\delta$  0.80, 0.84 and 1.01 were similar to those of the tertiary methyl groups in the molecule of *ent*-kaurane-16 $\alpha$ , 17-diol<sup>4</sup> isolated from *Fritillaria thunbergii* M, and the signals at chemical shifts of  $\delta$  0.84, 0.84, 0.99 were in

accord with those of the tertiary methyl groups in the molecule of *ent*-16 $\beta$ -hydroxy-kauran-17-oic acid<sup>5</sup>. The signals at chemical shifts of  $\delta$  4.02 and 3.86 belong to H-17' of **1**, but the corresponding signal of **2** is at  $\delta$  3.49 and 3.37. The hydroxyl of C-16' is tertiary and has stereoscopic obstruction, therefore, we concluded that the hydroxyl of C-17' is esterified. The <sup>13</sup>C-NMR spectrum showed 40 carbon signals (**Table 2**) which were assigned to one ester carbonyl carbon at  $\delta$  177.8 and 39 carbon signals at up-field.

Comparison of the <sup>1</sup>H-NMR and <sup>13</sup>CNMR chemical shifts of **1** with those of **2** and **3**, suggested that **1** is a dimer derived from **2** and **3**, Alkaline hydrolysis of **1** yielded two compounds **2** and **3** (Figure 1).



The structure of **2** and **3** were determined on the basis of spectroscopic and chemical evidence and by direct comparison with an authentic sample. Their spectral data were respectively identical with those reported in references (**Table 2**). The structure of **2**,  $C_{20}H_{34}O_2$  (EI-MS m/z 306 M<sup>+</sup>) was identified as *ent*-kaurane-16 $\alpha$ , 17-diol<sup>4</sup>. The structure of **3**,  $C_{20}H_{32}O_2$  (EI-MS m/z 304 M<sup>+</sup>) was identified as *ent*-16 $\beta$ -hydroxy kauran-17-oic acid<sup>5</sup>.

<b>1</b> δ (ppm)		<b>2</b> δ (ppm)		<b>3</b> δ (ppm)
0.80 (s, 3H, 18-CH <sub>3</sub> )		0.80 (s, 3H, 18-CH <sub>3</sub> )		0.80 (s, 3H, 18-CH <sub>3</sub> )
0.80 (s, 3H, 18'-CH <sub>3</sub> )		0.84 (s, 3H, 20-CH <sub>3</sub> )		0.85 (s, 3H, 20-CH <sub>3</sub> )
0.84 (s, 3H, 20-CH <sub>3</sub> )		1.03 (s, 3H, 19-CH <sub>3</sub> )		1.00 (s, 3H, 19-CH <sub>3</sub> )
0.84 (s, 3H, 20'-CH <sub>3</sub> )				
0.99 (s, 3H, 19-CH <sub>3</sub> )				
1.01 (s, 3H, 19'-CH <sub>3</sub> )				
4.02 (d, 1H, J=11.3Hz)		3.49 (d, 1H, J=11.3Hz)		
	-C-CH <sub>2</sub> -O-		-CH <sub>2</sub> OH	
3.86 (d, 1H, J=11.3Hz)		3.37 (1 d, H, J=11.3Hz)		

 Table 1
 <sup>1</sup>H-NMR spectral data for 1, 2, 3 (in CDCl<sub>3</sub>, TMS as inner standard, 300MHz)

Table 2 $^{13}$ C-NMR spectral data for 1, 2, 3, and ent-kaurane-16 $\alpha$ , 17diol, ent-16 $\beta$ -hydroxykauran-<br/>17-oic acid (300MHz, in CDCl<sub>3</sub>,  $\delta$  ppm)

С	1	2	3	ent-16β-hydroxykauran-17-oic	С	1	2	ent-kaurane-16a,17-diol	3
				acid					
1	41.6	4	1.8	39.2	1'	41.8	41.9	42.0	
2	18.6	1	8.6	18.2	2′	18.6	18.7	18.2	
3	42.0	4	2.0	42.0	3'	42.0	42.0	42.0	
4	33.2	3	3.2	33.1	4'	33.2	33.2	33.4	
5	56.0	5	6.1	56.1	5'	56.2	56.1	56.1	
6	20.7	2	20.4	20.6	6'	20.0	20.0	20.5	
7	38.3	3	9.8	40.3	7'	38.2	38.2	37.2	
8	45.6	4	5.3	45.3	8'	43.7	43.5	44.6	
9	57.0	5	7.0	56.0	9′	57.0	56.9	56.7	
10	39.3	3	9.2	38.0	10'	39.2	39.2	39.4	
11	18.4	1	8.0	18.5	11'	18.4	18.6	18.3	
12	26.7	2	27.4	31.2	12'	26.8	26.7	26.3	
13	45.1	4	4.4	44.7	13'	52.6	52.6	45.5	
14	40.4	4	0.7	40.8	14'	40.4	40.4	40.4	
15	45.0	4	5.3	45.0	15'	56.0	56.1	53.4	
16	56.1	5	6.0	55.9	16'	78.8	79.7	81.6	
17	177.8	1	80.4	182.5	17'	71.0	69.1	66.2	
18	33.5	3	3.6	33.5	18′	33.6	33.6	33.4	
19	21.6	2	21.5	21.5	19′	21.6	21.5	21.5	
20	17.4	1	7.6	17.3	20'	17.5	17.6	17.7	

From the above evidence, the structure of **1** was established as *ent*-16 $\alpha$ -hydroxy kauran-17-yl *ent*-16 $\beta$ -kauran-17-oate.

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